Phase 1 Clinical Trial of PSMA-directed/TGFβ-insensitive CAR-T cells in Metastatic Castration-Resistant Prostate Cancer: Safety, Correlative Studies, and Future Directions following Preliminary Dose Escalation

<u>Vivek Narayan, MD MSCE;</u>¹ Whitney Gladney, PhD;² Chris Kloss, PhD;² Gabriela Plesa, MD PhD;² Neha Vapiwala, MD;¹ Erica Carpenter, PhD;¹ Shannon Maude, MD PhD;^{2,3} Priti Lal, MD;¹ Simon Lacey, PhD;² J. Joseph Melenhorst, PhD;² Ronnie Sebro, MD PhD;¹ Michael Farwell, MD;¹ Wei-Ting Hwang, PhD;¹ Michael Moniak, RN BSN;² Joan Gilmore, BS CCRP;² Lester Lledo, CRNP;² Karen Dengel, RN BSN;² Carl June, MD;^{1,2} and Naomi Haas, MD.¹

- 1 Abramson Cancer Center, University of Pennsylvania, Philadelphia, PA
- 2 Center for Cellular Immunotherapies, University of Pennsylvania, Philadelphia, PA
- 3 Children's Hospital of Philadelphia, University of Pennsylvania, Philadelphia, PA

Background:

Adoptive immunotherapy with CAR-T cells has transformative potential for the treatment of cancer. However, a primary challenge to the success of these therapies in prostate cancer is the immunosuppressive microenvironment, including high levels of TGF β , encountered by re-directed T cells upon tumor infiltration. Importantly, these immunosuppressive functions of TGF β can be inhibited in T cells using a dominant negative TGF β receptor (TGF β Rdn), thereby enhancing antitumor immunity. In *in vivo* disseminated tumor models, co-expression of TGF β Rdn on PSMA-directed CAR-T cells led to increased T cell proliferation, enhanced cytokine secretion, long-term persistence, and greater induction of tumor eradication. Mechanisms of adaptive tumor resistance are unknown.

Methods:

We initiated a first-in-human phase 1 clinical trial to evaluate the safety and preliminary efficacy of lentivirally-transduced PSMA-directed/TGF β -insensitive CAR-T cells (CART-PSMA-TGF β Rdn) in men with treatment-refractory metastatic CRPC (NCT03089203). In preliminary dose-escalation cohorts, patients received a single dose of 1-3 x $10^7/m^2$ (Cohort 1) or 1-3 x $10^8/m^2$ (Cohort 2) CART-PSMA-TGF β Rdn cells without lymphodepleting chemotherapy in a 3+3 design. All treated patients underwent metastatic tumor biopsies at baseline, as well as on day +10 following the CAR-T cell infusion. Quantitative PCR of CART-PSMA-TGF β Rdn DNA was performed at serial timepoints to evaluate for CAR-T expansion and persistence in peripheral blood and trafficking to target tissues. Bioactivity of CART-PSMA-TGF β Rdn cells in peripheral blood was evaluated through Luminex analyses of immune and inflammatory factors.

Results:

Six patients have received CART-PSMA-TGFβRdn cell infusions at the specified dose levels (Cohort 1, N=3; Cohort 2, N=3). All CART-PSMA-TGFβRdn infusion products have met target transduction efficiency. Evaluation of CAR-T cellular kinetics via qPCR of CART-PSMA-TGFβRdn DNA has demonstrated peripheral blood T cell expansion (**Figure 1**), as well as tumor tissue trafficking in post-treatment tumor biopsies (**Table 1**). In Cohort 2, two patients developed anticipated Grade 3 cytokine release syndrome (CRS), and one patient developed Grade 3 CAR-T neurotoxicity requiring corticosteroids. Marked increases in inflammatory cytokines (IL-6, IL-15, IL-2, IFNgamma) and ferritin correlated with all Grade 3 CRS events (**Figure 2**). All CRS events rapidly resolved with tocilizumab (anti-IL6R) rescue. No dose-limiting toxicities have occurred.

Conclusions:

Cellular therapy with CART-PSMA-TGFRdn is safe and feasible in men with metastatic CRPC. CAR-T cell expansion and tumor tissue trafficking is observed. We observed clinically significant CRS, which is a critical marker of biologic activity with CAR-T therapy. In order to further

enhance CART-PSMA-TGF β Rdn expansion, persistence, and antitumor efficacy, ongoing cohorts are evaluating co-administration with lymphodepleting chemotherapy, as well as serial CART-PSMA-TGF β Rdn re-infusions. Critical planned analyses will examine mechanisms of adaptive resistance via comprehensive phenotyping of tumor-infiltrating T cells and the prostate cancer microenvironment, as well as circulating tumor material and T cell subtypes.

Conflict of Interest: None

Funding Acknowledgement: Prostate Cancer Foundation; Tmunity Therapeutics

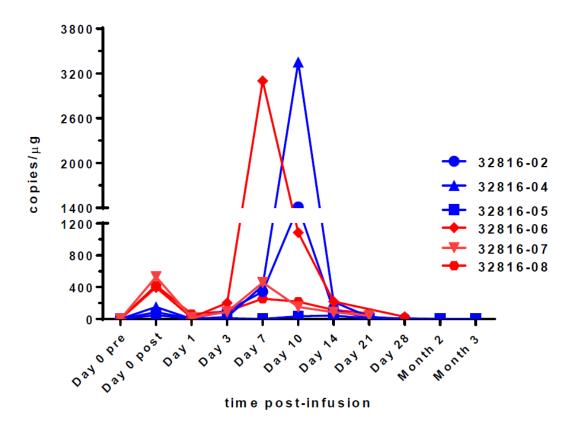


Figure 1. Detection of PSMA CAR by qPCR in the peripheral blood of prostate cancer subjects. Blue lines represent Cohort 1 subjects; red lines represent Cohort 2 subjects.

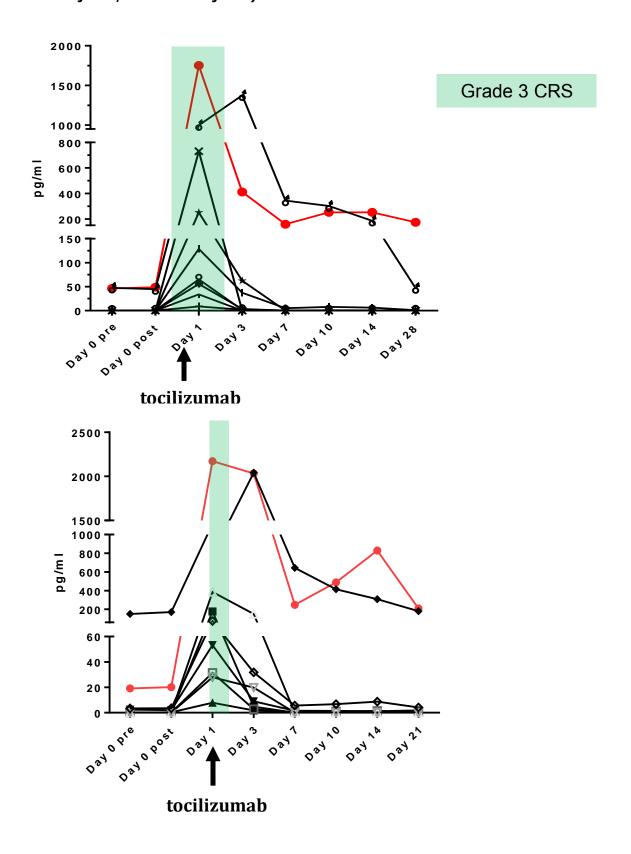
Table 1. CART-PSMA-TGFβRDN cell trafficking: qPCR detection in tissue biopsy samples in infused subjects

Subject ID	Cohort	Time point	Sample Type	Results*
32816-02	1	Day 10	Bladder (FFPE tissue curls)	122.32
			Bladder (FFPE tissue curls)	57.99
		Day 21	Bone marrow biopsy core	ND
			Bone marrow	27.12
		Month 2	Bone marrow biopsy core	ND
32816-04	1	Day 10	Bone (FFPE tissue curls)	133.36
			Bone (FFPE tissue curls)	211.16
32816-05	1	Day 10	Lymph node (FFPE tissue)	758.51
32816-06	2	Day 10	Lymph node (FFPE tissue)	98.24
32816-07	2	Day 10	Bone (FFPE tissue curls)	ND
32816-08	2	data analysis pending		

*copies/µg gDNA <u>Abbreviations</u> FFPE – formalin-fixed, paraffin embedded

ND – not detected

Figure 2. Inflammatory Cytokines in Subjects with Cytokine Release Syndrome (Top: Subject 6; Bottom: Subject 7)



- **→** IL10
- -▼- IL-7
- -A- IL-15
- -B- GM-CSF
- → IL2R
- **▼** IL2
- → TNFa
- **──** IFNgamma
- **→** IL6